FMPTS-25

Food Microbiology Proficiency Testing Scheme

Scheme Description

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Scheme Aims and Organization

The primary aim of the Food Microbiology proficiency testing scheme (FMPTS) is to enable laboratories performing the microbiological analysis of food to monitor their performance and compare it with that of their peers. FMPTS also aims to provide information to participants on technical issues and methodologies relating to testing of food.

The FMPTS scheme year operates from January to December. Further information about FMPTS, including test material availability, round dispatch dates and reporting deadlines, are available on the current contract/registration form.

Test Materials

Details of test materials available in FMPTS are given in Appendix A. The test parameters are continually reviewed to ensure they meet the needs of current laboratory testing and regulatory requirements.

Test materials are tested for homogeneity. Details of homogeneity tests performed and results are given in the FMPTS Scheme Reports and general protocol.

The planning of the scheme, the evaluation of performance and the authorization of the final report will never be subcontracted.

Statistical Analysis

Information on the statistics used in FMPTS can be found in the General Protocol and in the Scheme Report. Methods for determining assigned values and the values for SDPA used for individual samples are given in Appendix A

Methods

Participants will analyze PT sample depending on their routine method and the selection for the appropriate analysis method will be there responsibility.

Results and Reports

FMPTS reports will be available on the email within 14 working days of deadline. Participants will be emailed the report when it is available.

Reporting result

Results are reported on the form as a Correct number without Decimal places or Logs instead.

APPENDIX A - Description of abbreviations used

Assigned Value (AV)
The assigned value derive in the following ways:

- Consensus value from participant results: the use of a consensus value, produced in each round of the PT scheme, and based on the results obtained by the participants.

  The consensus value is usually estimated using robust statistical techniques (robust mean (R-Mean)). This is the mean of participant results after the removal of test results that are inappropriate for statistical evaluation (outlier > 5 Z-score), miscalculations, transpositions and other gross errors.

  Generally, the assigned value will be set using results from all methods, unless the measurement is considered method-dependent, in such case the assigned value will be set by method and indicated in the report tables.

The uncertainty of assigned values derived in this way is estimated from the participant results, according to ISO 13528:2022

- From a qualitative formulation (**Qual Form**). This applies to qualitative tests where the assigned value is simply based on the presence/absence of the analyte in the test material.

- **Range**: This indicates the concentration range at which the analyte may be present in the test material.

- **R-mean**: Robust mean

- **RSD**: Robust standard deviation.

- **SDPA**: standard deviation for proficiency assessment
  Which is used to assess participant performance for the measurement of each analyte This based on the robust standard deviation of the participant measurement results, either across all methods or by method depending on whether the measurement made is method dependent.
- **Units**: This indicates the units used for the assessment of data and in which participants should report their results. For some analytes in some schemes participants may have a choice of which units to report their results.
- **NA**: Not Applicable.
- **DP**: This indicates the number of decimal places to which participants should report their measurement results.
- **RR**: Result Rounding

Appendix A/ PT Items / Food /2025

Sample code: Sample PT-FM-101	Sample description: 1*10 ml vial glass (lyophilized) +1*10g milk powder		Instruction: to be resuscitated to final volume of 100ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Enumeration of Bacillus cereus	All	R-mean	0 – $1*10^5$	RSD	cfu /g	NA

Sample code: Sample PT-FM-100	Sample description: 1*10 ml glass vial (lyophilized) +1*10g milk powder		Instruction: to be resuscitated to final volume of 100ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Total aerobic mesophilic count	All	R-mean	0 – $1*10^5$	RSD	cfu /g	NA
Enumeration of coliforms	All	R-mean	0 – $1*10^5$	RSD	cfu /g	NA
Enumeration of Escherichia coli	All	R-mean	0 – $1*10^5$	RSD	cfu /g	NA
Enumeration of Enterobacteriaceae	All	R-mean	0 – $1*10^5$	RSD	cfu/g	NA

Sample code: Sample PT-FM-098	Sample description: 1*10 ml glass vial (lyophilized) +1*10g milk powder		Instruction: to be resuscitated to final volume of 100ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Enumeration of coagulase-positive Staphylococci	All	R-mean	0 – $1*10^5$	RSD	cfu/g	NA

Sample code: Sample PT-FM-097	Sample description: 1*10 ml glass vial (lyophilized) +1*10g milk powder		Instruction: to be resuscitated to final volume of 100ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Enumeration of yeast and mold	All	R-mean	0 – $1*10^5$	RSD	cfu/g	NA

Sample code: Sample PT-FM-096 (A,B)	Sample description: 2*10 ml glass vial (lyophilized) +1*50g milk powder		Instruction: to be resuscitated to final volume of 250ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Detection of Salmonella species	All	Qual Form	0 – $1*10^3$	NA	Present, absent/25g	NA

Sample code: Sample PT-FM-095	Sample description: 1*10 ml glass vial (lyophilized) +1*10g milk powder		Instruction: to be resuscitated to final volume of 100ml			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Enumeration of Clostridium perfringens	All	R-mean	0 – $1*10^5$	RSD	cfu/g	NA

Sample code: **Sample PT-FM-094 (A,B)**	**Sample description:** **2*10 ml glass vial (lyophilized)** **1*50g milk powder**		**Instruction:** **to be resuscitated to final volume of 250ml**			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Detection of Listeria monocytogenes	All	Qual Form	0 – $1*10^3$	NA	Present, Absent/25g	NA

Sample code: **Sample PT-FM-093 (A,B)**	**Sample description:** **2*10 ml glass vial(lyophilized)**		**Instruction:** **to be resuscitated to final volume of 250ml**			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP&RR**
Detection of Vibrio parahaemolyticus	All	Qual Form	0 – $1*10^3$	NA	Present, absent/25g	NA

Sample code: **Sample PT-FM-092 (A,B)**	**Sample description:** **2*10 ml glass vial (lyophilized)** **+1*50g milk powder**		**Instruction:** **to be resuscitated to final volume of 250ml**			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Detection of Escherichia coli O157	All	Qual Form	0 – $1*10^3$	NA	Present, absent/25g	NA

Sample code: **Sample PT-FM-091 (A,B)**	**Sample description:** **2*10 ml glass vial (lyophilized)**		**Instruction:** **to be resuscitated to final volume of 250ml**			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Detection of Campylobacter species	All	Qual Form	0 – $1*10^3$	NA	Present, absent/25g	NA

Sample code: **Sample PT-FM-090**	**Sample description:** **1*10 ml glass vial (lyophilized)** **+1*10g milk powder**		**Instruction:** **to be resuscitated to final volume of 100ml**			
Test	**Method**	**AV**	**Range**	**SDPA**	**Units**	**DP &RR**
Enumeration of listeria monocytogenes	All	R-mean	0 – $1*10^5$	RSD	cfu/g	NA